

Influence of Aqueous Solution of *Agaricus bisporus* (L) Treated Mulberry Leaves on the Quality of cocoons and silk filament in silkworm, *Bombyx mori* (L).

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Abstract

The aqueous solution of powder of fruiting body of *Agaricus bisporus* (L) with fifty milligram per liter (50 ppm) strength and aqueous solution of powder of AB21 protein with fifty milligram per liter (50 ppm) strength were used separately for treating mulberry leaves. Such treated mulberry leaves were fed daily to the silkworm larvae of bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27)] for first four days after the fourth moult. For each day, four feedings were supplied at the rate 100 grams of mulberry leaves for the group of hundred larvae. Larvae fed with untreated and water treated mulberry leaves were also maintained. Mature larvae were considered for the provision of moutage for spinning the cocoon. The cocoons were harvested on fifth day after the provision of moutage. The shell ratio of cocoons, denier scale of silk filament of the untreated control group; water treated group; AB21 protein treated group and fruiting body treated group were found measured 20.241, 2.734; 20.241, 2.734; 28.390, 3.006 and 33.309, 3.224 respectively. The contents of powder of fruiting body of *Agaricus bisporus* (L) including AB21 protein may be serving for enhancement of metabolism in the fifth larval instars of silkworm, *Bombyx mori* (L). Through the improved rate of metabolism, powder of fruiting body of *Agaricus bisporus* (L) and it's novel protein: "AB21-protein" treatment may aiming the action of gross metabolic constituency of silkworm larvae.

Keywords: *Bombyx mori* (L); *Agaricus bisporus* (L); AB21-protein; Shell Ratio

INTRODUCTION

Nutrition plays a pivotal role in sericulture by improving the commercial characters of silkworm. Silkworm being a monophagous insect derives almost all the nutrients required for its growth from the mulberry leaf itself (Nasreen *et al.*, 1999). Though the silkworm nutrients are balanced in mulberry leaf, the quantity available is not sufficient for the larval growth due to variation in mulberry plant cultivable soil (Ito, 1978). The intake of nutrient by the larvae is also proportional to the availability of feed. The

silkworm nutrition is considered as a major area of research in sericulture (Legay, 1958). Nutrition study on silkworm is an essential prerequisite for its proper commercial exploitation. Nutrition of silkworm is sole factor which almost individually augments quality and quantity of silkworm (Laskar and Datta, 2000). In recent year's attempts have been made in sericulture with nutrients such as proteins, carbohydrates, **amino acids**, vitamins hormones antibiotics etc. for better performance and to get high yield and quantity cocoons (Sannapa *et al.*, 2002; Etebari *et al.*, 2004). The salt significantly enhanced the growth of developmental stages and decreased the developmental period. Nickel chloride significantly increased the growth of larvae (Islam *et al.*, 2004). In addition to mulberry leaves feed supplements are also given to silkworm to enhance economic characteristics (Jeyapaul *et al.*, 2003; Sheeba *et al.*, 2006).

The dimeric protein AB21 from *Agaricus bisporus* (L), one of the most commonly and widely consumed mushrooms in the world. The protein shares no significant sequence similarity with any protein of known function, and it is the first characterized member of its protein family. The coding sequence of the "ab21" gene was determined and the protein was expressed in *E. coli* in a recombinant form. Komarek, *et al* (2018) demonstrated abundance of protein, entitled, "AB21" in the fruiting bodies of *Agaricus bisporus* (L). This protein deserve a high thermal and pH stability of AB21 and proved the weak affinity of the protein to divalent ions of some transition metals (nickel, zinc, cadmium, and cobalt). The reported crystallographic structure exhibits an interesting rod-like helical bundle fold with structural similarity to bacterial toxins of the ClyA superfamily.

In a 100-gram serving, raw white mushrooms provide 93 kilojoules (22 kilocalories) of food energy and are an excellent source (> 19% of the Daily Value, DV) of the B vitamins, riboflavin, niacin, and pantothenic acid (table). Fresh mushrooms are also a good source (10–19% DV) of the dietary mineral phosphorus (https://en.wikipedia.org/wiki/Agaricus_bisporus). While fresh *A. bisporus* only contains 0.2 micrograms (8 IU) of vitamin D as ergocalciferol (vitamin D₂), the ergocalciferol content increases substantially after exposure to UV light (Los Angeles Times, 2003 and Koyyalamudi, *et al*, 2009).

Mushrooms contain hydrazine derivatives, including agaritine and gyromitrin, that have been evaluated for carcinogenic activity (Hashida, *et al*, 1990). Agaritine, a hydrazine, poses no toxicological risk to humans when mushrooms are consumed in typical amounts (Roupasa, *et al*, 2010). No reports on use of edible mushroom fruiting bodies in sericulture.

The present study was therefore, undertaken to study the effect of aqueous solution of fruiting bodies of *Agaricus bisporus* (L) and aqueous solution of its novel protein: "AB21" on the quality of cocoons and silk filament in silkworm, *Bombyx mori* (L).

MATERIAL AND METHOD

The study was carried out through the steps like: Preparation of Aqueous Solution of fruiting bodies of *Agaricus bisporus* (L) Aqueous Solution of AB21-Protein; Rearing of silkworm larval stages [Race: (CSR6 x CSR26) x CSR2 x CSR27]; Treating the mulberry leaves and feeding larval instars; Analysis of characters of cocoons; silk filament and Analysis of the data through method of statistics.

(A). Preparation of Aqueous Solution of fruiting bodies of *Agaricus bisporus* (L) and Aqueous Solution of AB21-Protein:

The fruiting bodies of *Agaricus bisporus* (L) and the AB21-Protein were procured from Harry Organo Private Limited Ganpara, Durg - 491001, for Sericulture unit of Agricultural Development Trust,

Baramati (Malegaon Sheti Farm India). Addition of known volume of water was made in the bowl containing fruiting bodies of *Agaricus bisporus* (L) weigh of which was predetermined. This subjected for uniform mixing through the use of kitchen juice grinder for fifteen minutes. The content was filtered through the common laboratory filter paper. The filtrate was equalized with acetone to get the macerative of 50 ppm (mg/lit.) strength. In the similar manner, solution of AB21-Protein was prepared using water as a solvent. The strength of water solution of AB21-Protein was also 50 ppm (mg/lit.). Both were prepared freshly before using for treating the mulberry leaves.

(B). Rearing of silkworm larval stages [Race: (CSR6 x CSR26) x CSR2 x CSR27]: The method of Krishnaswami, *et al*, (1992) for rearing of silkworm larvae was followed. This method is appearing in the document authorized by Khyade (2004) and Vitthalrao B. Khyade, *et al* (2005, 2006, 2007, 2008, 2009, 2010, 2011, 2012, 2013, 2014, 2015, 2016, and 2017).

The disease free layings of bivoltine race (CSR6 x CSR26) x CSR2 x CSR27) of silkworm, *Bombyx mori* (L) were procured through the “Dr. APIS” Laboratory and processed for black boxing, rearing of early instars, rearing of late age instars, provision of moutage for spinning the cocoon and cocoon harvesting through the standard methods.

(C). Treating the mulberry leaves and feeding larval instars: The fifth instar larvae were utilized for the carrying out the attempt. Soon after the fourth moult, the fifth instared larvae were grouped into four groups (each with hundred individuals). The groups include:

1. The Group of Untreated Control;
2. The Group of Water treated control;
3. Group of larvae fed with mulberry leaves treated with aqueous solution of fruiting bodies of *Agaricus bisporus* (L) and
4. Group of larvae fed with mulberry leaves treated with Aqueous Solution of AB21-Protein.

For treatment, known quantity of mulberry leaves was kept immersed in known volume of aqueous solution (at the rate hundred grams of mulberry leaves in liter of aqueous solution) for half an hour. Such treated mulberry leaves were fed daily to the silkworm larvae of bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27)] for first four days after the fourth moult. For each day, four feedings were supplied at the rate 100 grams of mulberry leaves for the group of hundred larvae. The larvae of the group of Untreated Control were received untreated mulberry leaves. The larvae of the group of Water Treated Control were received water treated mulberry leaves. From sixth day onwards, the larvae of all the groups were fed with untreated mulberry leaves through standard methods. Rearing was carried out in the trays of wood. For each day, larvae received four feedings (at the rate of hundred grams of mulberry leaves for the group of hundred larvae for each feeding). The moutage was made available for spinning the silky cocoon by the mature last larval stages Khyade, 2004 and Vitthalrao B. Khyade, *et al* (2005, 2006, 2007, 2008, 2009, 2010, 2011, 2012, 2013, 2014, 2015, 2016, 2017 and 2018).

(D). Analysis of economic parameters: The cocoons were separated from the moutage. Separation of cocoons from moutages is called as harvesting. This harvesting of cocoons was carried out on fifth day after the provision of moutage for spinning. Twenty cocoons from each group were selected randomly. They were deflossed. The weight of individual deflossed cocoon was recorded. Each cocoon in the was cut vertically using the blade and weight of pupa was recorded. For knowing the shell weight of individual cocoon, the reading of the weight of pupa was subtracted from weight of respective cocoon. Weight of entire deflossed cocoon; weight of shell of cocoon and weight of pupa were noted. The silk shell percentage (correctly called as shell ratio) was calculated through the use of readings of weight of whole deflossed cocoon and weight of silk shell in cocoon. The mathematical operation of dividing the

readings of silk shell weight by readings of weight of whole cocoon without floss was followed. Multiplication operation was carried with hundred with quotient obtained earlier. This yields the shell percentage. In sericulture, this silk shell percentage is called as shell ratio.

Ten cocoons per replication were used for the purpose to reel the silk filament from individual cocoon. The length in meter (A) of unbroken silk filament was obtained by using eprouvate. Weight in gram of silk filament (B) from individual cocoon was recorded. Length (A) and weight (B) of silk filament were accounted for the calculation of Denier scale. The reading of weight of silk filament (B) was divided by the reading of length of silk filament (A). Quotient thus obtained was multiplied by 9000 for the purpose to get the denier scale of silk filament Vitthalrao B. Khyade and Abhilasha C. Bhunje, 20015 and 2016).

(E). Statistical Analysis of the data:

The experimentation was replicated for three times. This is for the purpose to get the consistent results. The data was collected and it was subjected for statistical analysis. The statistical parameters considered in the attempt include: mean, standard deviation, percent variation and student “t” - test (Norman and Bailey,1955).

RESULTS AND DISCUSSION

The results on the attempt on the study of influence of aqueous solution of *Agaricus bisporus* (L) treated mulberry leaves on the quality of cocoons and silk filament in silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] are summarized in table-1 and presented in Fig. 1 and 2. The weight (gm) of entire deflossed cocoon; weight of silk shell (gm) and the weight of Pupa (gm) of the Untreated Control group were measured 1.823 (± 0.088); 0.369 (± 0.013) and 1.454 respectively. The ratio of shell to the entire cocoon in the untreated control group was calculated 20.241. The readings 796.47 (± 9.616); 0.242 (± 0.049) and 2.734 belongs respectively to the Silk Filament Length (SFL in meters); Silk Filament Weight (SFW in grams) and the denier scale of silk filament obtained from the untreated control group cocoons (Table – 1 and Fig. 1 and 2).

The weight of whole cocoon (deflossed), silk shell weight, weight of pupa, silk shell percentage or ratio and denier scale (unit of measurement of quality of filament) of the water treated group was found measured 1.823 (± 0.095); 0.369 (± 0.014); 1.453; 20.241 and 2.734 respectively (Table – 1 and Fig. 1 and 2).

First four days feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] with leaves of mulberry, *Morus alba* (L) treated with aqueous solution of fruiting bodies of *Agaricus bisporus* (L) was found resulted into spinning the cocoons weighing 2.684 (± 0.235).

The weight of pupa; weight of shell and shell ratio of cocoons of this group (larvae received leaves of mulberry, *Morus alba* (L) treated with aqueous solution of fruiting bodies of *Agaricus bisporus* L) was found measured 1.922 gm; 0.762 gm (± 0.067) and 28.390 respectively.

The length, weight and denier scale of silk filament reeled from cocoons of of this group (larvae received leaves of mulberry, *Morus alba* (L) treated with aqueous solution of fruiting bodies of *Agaricus bisporus* L) was found measured 1008.77 (± 58.924); 0.337 gm (± 21.786) and 3.006 respectively.

Feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] (for the first four days) with mulberry leaves treated with aqueous solution of AB21 protein was found resulted into spinning the cocoons weighing 2.756 (± 0.317).

The weight of pupa; weight of shell and shell ratio of cocoons of this group (larvae received leaves of mulberry, *Morus alba* (L) treated with aqueous solution of AB21 protein) was found measured 1.838 gm; 0.918 gm (± 0.119) and 33.309 respectively. First four days feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] with leaves of mulberry, *Morus alba* (L) treated with aqueous solution of fruiting bodies of *Agaricus bisporus* (L) was found resulted into spinning the cocoons weighing 2.684 (± 0.235).

The weight of pupa; weight of shell and shell ratio of cocoons of this group (larvae received leaves of mulberry, *Morus alba* (L) treated with aqueous solution of fruiting bodies of *Agaricus bisporus* L) was found measured 1.922 gm; 0.762 gm (± 0.067) and 28.390 respectively.

The length, weight and denier scale of silk filament reeled from cocoons of of this group (larvae received leaves of mulberry, *Morus alba* (L) treated with aqueous solution of fruiting bodies of *Agaricus bisporus* L) was found measured 1008.77 (± 58.924); 0.337 gm (± 21.786) and 3.006 respectively.

Feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] (for the first four days) with mulberry leaves treated with aqueous solution of AB21 protein was found resulted into spinning the cocoons weighing 2.756 (± 0.317).

The weight of pupa; weight of shell and shell ratio of cocoons of this group (larvae received leaves of mulberry, *Morus alba* (L) treated with aqueous solution of AB21 protein) was found measured 1.838 gm; 0.918 gm (± 0.119) and 33.309 respectively.

Table-1: The quality of the cocoons and silk filament spun by mature fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] separately received mulberry leaves treated with aqueous Solution of *Agaricus bisporus* (L) and aqueous solution of dimeric protein “ab21” a novel mushroom protein.

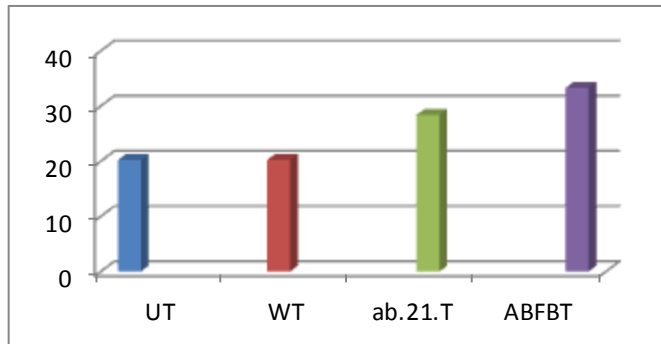
Parameters→ Group↓	Cocoon Weight (gm)	Shell Weight (gm)	Pupal Weight (gm)	Shell Ratio	S F L (m) (A)	S F W (gm) (B)	Denier Scale of S F = (B÷A) x 9000
Untreated Control (UT)	1.823 (±0.088) 00.000	0.369 (±0.0013) 00.000	1.454 00.000	20.241 00.000	796.47 (±9.616) 00.000	0.242 (±0.049) 00.000	2.734 00.000
Water Treated Control (WTC)	1.823 (±0.095) 00.000	0.369 (±0.014) 00.000	1.454 00.000	20.241 00.000	796.47 (±13.788) 00.000	0.242 (±0.055) 00.000	2.734 00.000
Treated with aqueous solution of dimeric protein “ab21” (ab.21.T)	2.684** (±0.235) 47.229	0.762** (±0.067) 106.50	1.922* 32.187	28.390** 8.149	1008.77* (±58.924) 26.655	0.337** (±21.786) 29.752	3.006** 09.948
Treated with aqueous solution of Fruiting Body of <i>Agaricus bisporus</i> (ABFBT)	2.756*** (±0.317) 51.179	0.918*** (±0.119) 148.780	1.838*** 26.409	33.309*** 13.068	1141.47*** (±97.858) 43.316	0.409*** (±0.091) 69.008	3.224*** 17.922

- Each figure is the mean of the three replications.

-Figure with ± sign in the bracket is standard deviation.

-Figure below the standard deviation is the increase for calculated parameter and percent increase for the others over the control. UTC=Untreated Control; WTC = WaterTreated Control; ab.21.T: ab.21 Treated; ABFBT: Treated with aqueous solution of *Agaricus bisporus* Fruiting Body; SFL= Silk Filament Length; SFW= Silk Filament Weight; *: P < 0.05; **: P < 0.005; ***: P < 0.01

Fig. 1: The Shell Ratio of the cocoons spinned by mature fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] separately received mulberry leaves treated with aqueous Solution of *Agaricus bisporus* (L) and aqueous solution of dimeric protein “ab21” a novel mushroom protein.



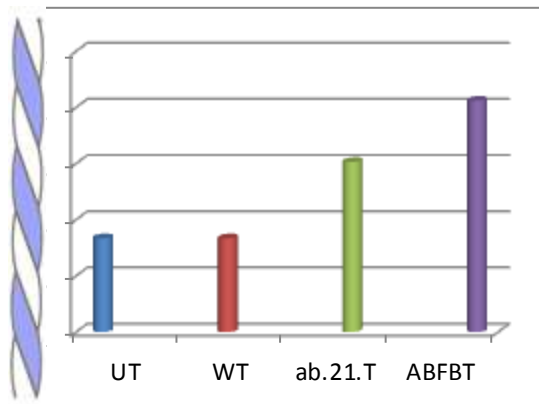
UT=Untreated Control;

WT =Water Treated Control;

ab.21.T: ab-21 Protein Treated;

ABFBT: Treated with water solution of *Agaricus bisporus*

Fig. 2: The Denier Scale of Silk Filament Reeled from the cocoons spinned by mature fifth instar larvae of silkworm, *Bombyx mori* (L) [Race: (CSR6 x CSR26) x CSR2 x CSR27)] separately received mulberry leaves treated with aqueous Solution of *Agaricus bisporus* (L) and aqueous solution of dimeric protein “ab21” a novel mushroom protein.



UT=Untreated Control;

WT =Water Treated Control;

ab.21.T: ab-21 Protein Treated;

ABFBT: Treated with water solution of *Agaricus bisporus*

The silk cocoon is foremost feature in sericultural practices. This is because, silkworm cocoons are sole source for silk commercially available. Silk cocoons are used to obtain the silk filament. The weight of cocoon weight; weight of silk shell and therein the silk shell percentage or shell ratio in present attempt of

study were found effected through treating the mulberry leaves with aqueous solution of fruiting bodies of *Agaricus bisporus* (L) and aqueous solution of AB21 protein feeding separately to the fifth instar larvae of silkworm. The mathematical range of increase in percentage of the cocoon weight and silk shell weight in the experimental (treated) groups was 47.229 to 51.179 and 106.50 to 148.780 respectively.

Silk shell percentage or shell ratio of the cocoons was found improved in the corresponding groups of treatment. Both, AB21 Protein, a known and novel mushroom protein and agaricus fruiting body, used for treating mulberry leaves before feeding fifth instar larvae in the present attempt were found suggestively the most significant ($p < 0.001$) with reference to the yield of the silk cocoons through silk shell percentage or shell ratio.

The growth and development of silkworm is under the continuous influence of factors operating within and outside the body (Murugan *et al.*, 1998). Ascorbic acid had effect on the growth of silkworm (Javed and Gondal, 2002) and combination of 0.2% of N which enhances the growth of silk production (Hussain and Javed, 2002). It is evident from the mean data of the experiments that, AB21-Protein treated leaves fed larvae showed a significant enhancement in reeling performance and bioenergetics. Sarkar *et al.* (1995) reported that growth of larvae *B.mori* significantly improved when they were fed on mulberry leaves supplemented with different nutrients such as Soya milk, Milk powder, Sugars, vitamins and amino acids. Rate of feeding influences the synthesis of total DNA, RNA and protein synthesis (Chavancy and Fournier, 1979). According to Soo-Hoo and Frankel (1966) the diminishing consumption rate of less preferred food was partially compensated by increased assimilation efficiency. However, according to Mathavan and Krishnan (1976) assimilation efficiency did not vary significantly as a function of reduced food consumption. Verma and Atwal (1963) observed that feeding leaves supplemented with distilled water alone slightly increased the weights of larva, pupa and silk shells. The significant results obtained in the attempt may be through the integrative action of the contents of fruiting bodies of *Agaricus bisporus* (L) and it's novel protein: AB21. This attempt on the use of AB21 Protein and the agaricus fruiting bodies through water for treating mulberry leaves before feeding silkworm larvae is much more easy method and may open a new avenue in the sericulture.

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REFERENCES

1. Ayoola, G. A. (2008). "Phytochemical Screening and Antioxidant Activities of Some Selected Medicinal Plants Used for Malaria Therapy in Southwestern Nigeria". *Tropical Journal of Pharmaceutical Research*. 7 (3): 1019–1024. doi:10.4314/tjpr.v7i3.14686.
2. Chavancy, G. and A. Fournier, 1979. Effect of starvation on t-RNA synthesis amino acid pool-RNA synthesis activities in the posterior silk gland of *Bombyx mori* L. *Biochimie.*, 61: 229-243.PubMed
3. Etebari, K., R. Ebadi and L. Matindoost, 2004. Effect of feeding mulberry enriched leaves with ascorbic acid on some biological, biochemical and economical characteristics of silkworm *Bombyx mori* L. *Int. J. Entomol.*, 8: 81-877.
4. Hashida C, Hayashi K, Jie L, Haga S, Sakurai M, Shimizu H (June 1990). "[Quantities of agaritine in mushrooms (*Agaricus bisporus*) and the carcinogenicity of mushroom methanol

- extracts on the mouse bladder epithelium]". *Nippon Koshu Eisei Zasshi (in Japanese)*. **37** (6): 400–5. PMID 2132000.
5. https://en.wikipedia.org/wiki/Agaricus_bisporus
 6. Hussain, M. and H. Javed, 2002. Effect of 0.2% of N with various combination of ascorbic acid on growth and silk production of silkworm *Bombyx mori* L. *Asian J. Plant Sci.*, 1: 650-651.
 7. Islam, M.R., A.O. Ali, D.K. Paul, S. Sultana, N.A. Banu and M.R. Islam, 2004. Effect of salt Nickel chloride supplementation on the growth of silkworm *Bombyx mori* L (Lepidoptera: Bombycidae). *J. Biol. Sci.*, 4: 170-172.
 8. Ito, T., 1978. Ascorbic acid is reported to the host plant mulberry morus Indica. L. *Indian J. Expt. Bio.*, 4: 31-36.
 9. Javed, H. and M.H. Gondal, 2002. Effect of food supplementation by n and ascorbic acid on larval mortality of silkworm (*Bombyx mori* L.). *Asian J. Plant Sci.*, 1: 556-557.
 10. Jeyapaul, C., C. Padmalatha, and A.J.A. Ranjith Singh, 2003. Effect of plant extracts on nutritional efficiency in mulberry silkworm, *Bombyx mori* L. *Indian J. Seric.*, 42: 128-131.
 11. Khyade V. B.; Gaikwad D. R. and Thakare U. G. (2012). Utilization of *Aloe vera* (L) Herbal Tonic for Treating Mulberry Leaves before feeding the Fifth Instar Larvae of Silkworm, *Bombyx mori*(L) (*Race: PM x CSR2*) (Editor: Dr. A. R. Tuwar and Dr. M. J. Shaikh Dept. of Life Sciences, Arts and Science College, Sonai Tal. Newasa, Dist. Ahmednagar – 414105 India): 37 – 40.
 12. Khyade, V. B. (2004). Influence of juvenoids on silk worm, *Bombyx mori* (L). Ph.D. Thesis, Shivaji University, Kolhapur, India, 2004.
 13. Khyade, V. B.; Patil, S. B.; Khyade, S. V. and Bhawane, G. P. (2002). Influence of acetone maceratives of *Vitis vinifera* on the larval parameters of silk worm, *Bombyx mori* (L), *Indian Journal of Comparative Animal Physiology*, 2002, 20, 14 -18.
 14. Khyade, V. B.; Ganga V. Mhamane (2005). Vividh Vanaspati Arkancha Tuti Reshim Kitak Sangopanasathi Upyojana, *Krishi Vdnyan*, 2005, 4, 18-22.
 15. Khyade, V. B.; Patil, S. B.; Khyade, S. V. and Bhawane, G. P. (2003). Influence of acetone maceratives of *Vitis vinifera* on the economic parameters of silk worm, *Bombyx mori* (L). *Indian Journal of Comparative Animal Physiology*, 2003, 21, 28–32.
 16. Khyade, V. B.; Poonam B. Patil; M. Jaybhay; Rasika R. Gaikwad; Ghantaloo, U. S.; Vandana D. Shinde; Kavita H. Nimbalkar and J. P. Sarwade (2007). Use of digoxin for improvement of economic parameters in silk worm, *Bombyx mori* (L). *Bioinformatics (Zoological Society of India)*, 2007.
 17. Khyade, V. B.; Sonali S. Machale; J. P. Sarwade; S. B. Patil and Sadhana H. Deshpande (2006). Screening of plant extractives for juvenoid activity in silk worm, *Bombyx mori* (L). *Journal of Zoological Society of India: Environment and Development*: 61 – 77.(Editors: B. N. Pandey and G. K. Kulkarni) (Publisher: A P H Publishing Corporation, New Delhi) (ISBN: 81-313-004-8 / 97881315300497).
 18. Komarek, J.; Ivanov Kavkov, E.; Houser, J.; Horackova, A.; Zdanska, J.; Demo, G. and Wimmerova, M. (2018). Structure and properties of AB21, a novel *Agaricus bisporus* protein with structural relation to bacterial pore-forming toxins. *Proteins*. 2018 May 2. doi: 10.1002/prot.25522. <https://www.ncbi.nlm.nih.gov/pubmed/29722060>.
 19. Koyyalamudi SR, Jeong SC, Song CH, Cho KY, Pang G (April 2009). "Vitamin D2 formation and bioavailability from *Agaricus bisporus* button mushrooms treated with ultraviolet irradiation". *Journal of Agricultural and Food Chemistry*. **57** (8): 3351–5. doi:10.1021/jf803908q. PMID 19281276.

20. Krishnaswami, S.; Narasimhana, M. N.; Suryanarayana, S. K. and Kumaraj, S. (1978). Sericulture Manual –II: Silk worm rearing. F A O, United Nation's Rome, 1978, 131.
21. Krishnaswami, S., M.N. Natrasimhana, S.K. Suryanarayanan and S. Kumaraj, 1973. Manual on Sericulture. Food and Agriculture Organisation, Rome, Italy.
22. Kumar, C.S., A.K. Goel, S.V. Seshagiri, S.S. Kumari, H. Lakshmi, C. Ramesha and C.M. Anuradha, 2009. Nutrigenetic traits analysis for the identification of nutritionally efficient silkworm germplasm breeds. *Biotechnol.*, 9: 131-141.
23. Laskar, N. and M. Datta, 2000. Effect of alfalfa tonic and its inorganic ingredients on growth and development of silkworm *Bombyx mori* L. race Nistari. *Environ. Ecol.*, 18: 591-596.
24. Laszczyk, Melanie (2009). "Pentacyclic Triterpenes of the Lupane, Oleanane and Ursane Group as Tools in Cancer Therapy". *Planta Medica*. 75 (15): 1549–60. doi:10.1055/s-0029-1186102. PMID 19742422.
25. Legay, J.M., 1958. Recent advances in silkworm nutrition. *Ann. Rev. Entomol.*, 3: 75-86.
26. Mathavan, S. and J.M. Krishnan, 1976. Effects of ration levels and restriction of feeding durations on food utilization in *Danaus chrysippus* (Lepidoptera: Daniadae). *Entomol. Exp. Appl.*, 19: 155-162.
27. Murugan, K., D. Jeyabalan, N. Senthil Kumar, R. Babu, N. Sivapirakasam and S.S. Nathan, 1998. Growth promoting effects of plant products on silkworm. A biotechnology approach. *J. Sci. Indian Res.*, 57: 740-74.
28. Mushrooms and vitamin D. Los Angeles Times. Retrieved 23 August 2003.
29. Nasreen, A., G.M. Cheema and M. Ashfaq, 1999. Rearing of silkworm *Bombyx mori* L. on alternative food parts. *Pak. J. Biol. Sci.*, 2: 843-845.
30. Roupasa P, Keogh J, Noakes M, Margettsa C, Taylor P (April 2010). "Mushrooms and agaritine: A mini-review". *Journal of Functional Foods*. 2 (2): 91–8. doi:10.1016/j.jff.2010.04.003.
31. Sannapa, B., M.J. Ramaiah and D. Chandrappa, 2002. Influence of castor genotype on consumption indices of eri silkworm sumia *Cynthia ricini*. *Bioduval. Environ. Ecol.*, 20: 960-964.
32. Sarkar, A., M. Rab and N. Absar, 1995. Effects of feeding mulberry (*Morus* sp).Leaves supplemented with different nutrient to silkworm (*Bombyx mori*) L. *Curr. Sci.*, 69: 185-188.
33. Sengupta, K., B.D. Singh and C. Mustafij, 1972. Nutrition of Silkworm. *Bombyx mori* L. I. Studies on the enrichment of mulberry leaf with various sugars, proteins, amino acids and vitamin for vigorous growth of the worm and increased cocoon crop production. *Indian J. Sci.*, 11: 11-27.
34. Sheeba, D.V., C. Padmalatha and A.J.A.R. Singh, 2006. Effects of supplementation of aminoacid, leucine and valine on the economic characters of silkworm. *J. Zool.*, 26: 277-280.
35. Soo-Hoo, C.F. and G. Frankel, 1966. The consumption, digestion and utilization of food plants by a poly phagous insect, *Prodenia eridania* (cramer). *J. Insect. Physiol.*, 12: 711-730.
36. Verma, A.N. and A.S. Atwal, 1963. Effect of chloromycetin and molasses on the growth and production of silk by *Bombyx mori* L. (Lepidoptera:Bombycidae). *Indian J. Seric.*, 1: 1-14.
37. Vishakha S. Chape; Abhilasha C. Bhunje and Vitthalrao B. Khyade (2016). Efficient Use of Extractive of *Oroxylum indicum* for the improvement of Quality of Silk in Silkworm, *Bombyx mori* (L) (Race: PM x CSR2). International Conference on "Plant Research and Resource

- Management” And 25th APSI Silver Jubilee Meet 2016 at T. C. College Baramati 11, 12 and 13 February, 2016. Pages: 304 – 308.
38. Vitthalrao B Khyade and Vivekanand V Khyade (2013). The Phytocompounds of Animal Hormone Analogues. Annals of Plant Sciences Vol. 2 (5): 125 – 137. <http://annalsofplantsciences.com/index.php/aps/issue/view/10> ISSN: 2287 – 688X
 39. Vitthalrao B. Khyade (2014): Influence of Lanoxin Treated Mulberry Leaves on the contents of proteins in the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR2). 2014. (Page: 8 – 17). Proceeding, Two day UGC sponsored National seminar on, “Recent Trends in Cell Biology, Biotechnology and Bioinformatics”, Organized by Department of Zoology, Balwant College, Vita Tal. Khanapur, Dist. Sangli 415311 (India) (6 and 7 September, 2013). Editor: Prof. (Smt.) U. H. Shah (Department of Zoology, Balwant College, Vita). ISBN 978 – 81 – 927211 – 3 – 2.
 40. Vitthalrao B. Khyade (2005). Vividh Vanaspati Arkancha Tuti Reshim Kitak Sangopanasathi Upyojan. Influence of mealy bug infestation on mulberry leaves on the silkworm, *Bombyx mori*(L). Krishi Vidnyan 4: 18 – 22.
 41. Vitthalrao B. Khyade (2014).THE ACTIVITY OF PROTEASE IN THE FIFTH INSTAR SILKWORM, *BOMBYX MORI* (L) (RACE: PM X CSR2). Biolife April – June Vol. 2 (2) 2014:
 42. Vitthalrao B. Khyade (2016). The Pattern of Chitin Deposition in the Integument of Fifth Instar Larvae of Silkworm for Topical Application of Acetone Solution of Triterpene Compounds . International Academic Journal of Innovative Research Vol. 3, No. 10, 2016, pp. 1-31.ISSN 2454-390X <http://iaiest.com/dl/journals/8-%20IAJ%20of%20Innovative%20Research/v3-i10-oct2016/paper1.pdf>
 43. Vitthalrao B. Khyade (2016). Utilization of mulberry leaves treated with seed powder of cowpea, *Vigna unguiculata* (L) for feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR2). Journal of Medicinal Plants Studies 2016; 4(3): 182 - 188. <http://www.plantsjournal.com/archives/2016/vol4issue3/PartC/4--2-33-339.pdf>
 44. Vitthalrao B. Khyade; Sunanda V. Khyade; Vivekanand V. Khyade; Sharad G. Jagtap and Jeevan P. Sarwade (2009). Tyrosine aminotransferase in the silkworm, *Bombyx mori*(L) (Race: PM x CSR₂). Advances in Pollution Research. Vol.21 (1): 1 – 4.
 45. Vitthalrao B. Khyade and Abhilasha C. Bhunje (2015).Efficient use of acetone extractive of *Oroxylum indicum* for the improvement of quality of silk in silkworm *Bombyx mori* (L.) (Race: PM x CSR2). Malaya Journal of Biosciences 2015, 2(4):185-190 ISSN 2348-6236 print /2348-3075 online <http://www.malayabiosciences.com/>
 46. Vitthalrao B. Khyade and Anil N. Shendage (2012). Influence of *Aloe vera* (L) Herbal formulation on Larval Characters and Economic Parameters of silkworm, *Bombyx mori* (L)(Race: PM x CSR2). The Ecoscan Special Issue Vol. 1 (121): 321 – 326. www.theecoscan.in ISSN: 0974 – 0376.
 47. Vitthalrao B. Khyade and Atharv Atul Gosavi (2016).Utilization of mulberry leaves treated with seed powder cowpea, *Vigna unguiculata* (L) for feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR2). World Scientific news 40 (2016): 147-162. www.worldscientificnews.com .
 48. Vitthalrao B. Khyade and Dhanashri R. Gaikawad (2016). Insect Juvenile Hormone. World Scientific News 44 (2016): 216-239. www.worldscientificnews.com.
 49. Vitthalrao B. Khyade and Jivan P. Sarwade (2009). Influence of methanolic extractives of roots of *Achyranthus aspera* (L) on the body wall chitin in fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR₂). Journal of Association of Zoologists, India. Vol. 2 (1): 11 – 21.

50. Vitthalrao B. Khyade and Jiwan P. Sarwade (2009). Influence of acetone extractives selected plants on the body wall chitin of fifth instars of silkworm, *Bombyx mori* (L) (Race: PM x CSR₂). Journal of Association of Zoologists, India. Vol. 2 (1): 39 – 47.
51. Vitthalrao B. Khyade and Jiwan P. Sarwade (2009). Protein profiles in the fifth instar larvae of silkworm, *Bombyx mori*(L) (Race: PM x CSR₂), fed with Digoxin treated mulberry leaves. The Bioscan, Vol.4, No.1: 41 – 44.
52. Vitthalrao B. Khyade and Jiwan P. Sarwade (2013): Utilization of Digoxin, the herbal product for treating the mulberry leaves and feeding the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR₂). 2013 International Journal of Multidisciplinary Research (IJMR) Vol. I / Issue 12 (III): 38-42. ISSN: 2277 – 9302.
53. Vitthalrao B. Khyade and Jiwan P. Sarwade (2013): Utilization of Retinol through the topical application to the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR₂) for qualitative improvement of the economic parameters. International Journal of Advance Life Sciences Vol. 6 Issue 5 November, 2013. Pages: 532 – 537. www.ijals.com http://ijals.com/wp-content/uploads/2014/01/19.-Utilisation-of-Retinol-through-the-topical.pdf
54. Vitthalrao B. Khyade and Jyoti A. Kulkarni (2011). Effect of Digoxin treated mulberry leaves on protein profiles in fifth instar larvae of Silkworm, *Bombyx mori*(L) (PM x CSR₂). Research Journal of Chemical Sciences Vol.1 (1): 2 – 6. www.isca. ISSN 2231.
55. Vitthalrao B. Khyade and K. Slama (2014). Changes in the Pattern of Chitin Deposition in The Integument of Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (Pm X Csr₂) Topically Applied With Various Concentrations Of Acetone Solution Of Retinol. Journal of Biodiversity and Ecological Sciences Vol. 4, Issue 4: 159 – 167. ISSN: 2008-9287.
56. Vitthalrao B. Khyade and Karel Slama (2015). SCREENING OF ACETONE SOLUTION OF FME AND SELECTED MONOTERPENE COMPOUNDS FOR JUVENILE HORMONE ACTIVITY THROUGH CHANGES IN PATTERN OF CHITIN DEPOSITION IN THE INTEGUMENT OF FIFTH INSTAR LARVAE OF SILKWORM, *Bombyx mori* (L) (PM x CSR₂). IJBRTISH Vol. 2 Issue 3 (May – June 2015): 68 – 90. ISSN 2349-9419 www.ijbritish.com
57. Vitthalrao B. Khyade and M. B. Deshmukh (2004). Evaluation of plant extracts for juvenoid activity against red cotton bug, *Dysdercus cingulatus* (L). Influence of mealy bug infestation on mulberry leaves on the silkworm, *Bombyx mori*(L). The Proceeding of International Symposium (23 – 25 November, 2004); University of Agricultural Sciences, Dharwad, Karnataka (India) on strategies for sustainable cotton production: A global vision/ 3. crop protection: 97 – 99.
58. Vitthalrao B. Khyade and Rajkumar B. Deshmukh (2015). Mid gut protease and amylase activity in the fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR₂) fed with mulberry leaves treated with aqueous solution of stevia inulin powder. Proceedings, U G C Sponsored National Conference on Recent Trends in Life Sciences (10 - 11, July, 2015), organized by Department of Zoology, S. M. Joshi College, Pune. Page: 95 – 106. ISBN 978-93-5235-362-0.
59. Vitthalrao B. Khyade and Sucheta S. Doshi (2012). Protein Contents and activity of enzymes in the mid gut homogenate of fifth instar larvae of silk worm, *Bombyx mori*(L) (Race: PM x CSR₂) fed with herbal drug (Kho Go) treated mulberry leaves. Research Journal of Recent Sciences Vol. 1 (2): 49 – 55. www.isca.in ISSN 2227 – 2502.
60. Vitthalrao B. Khyade and Vivekanand V. Khyade (2013): Plants: The Source of Animal Hormones. “Frontiers in Life sciences”, the book published by Science Impact Publication, Ahmedpur (Latur) – 413515 (India): 151 – 168. Editor: Dr. Sayyed Iliyas Usman (Poona College, Camp Pune). ISBN: 978 – 93 – 5067 – 394 – 2.

61. Vitthalrao B. Khyade, Kajal D. Gokule, Sunanda Rajendra Pawar, Rajkumar B. Deshmukh (2016). Utilization of the Retinol and Phytol for the quality improvement of cocoon and silk fibre spinned by fifth instar larvae of silkworm, *Bombyx mori* (L) (Race: PM x CSR2). World Scientific News 42 (2016): 167-181. www.worldscientificnews.com.
62. Vitthalrao B. Khyade, Sivani C. Bhosale; Vishakha R. Kakade and Jiwan P. Sarawade (2015). Pattern of Chitin Deposition in The Integument of Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (PM x CSR2) Treated with Acetone Solution of Selected Monoterpene Compounds and Farnesol Methyl Ether (Fme). Journal of Basic Sciences, 2015, Special Issue on BioIPPF, 34-40. www.skpubs.com
63. Vitthalrao B. Khyade, Vivekanand V. Khyade and Amar H. Kadare (2014): Influence of Acetone Extractive of *Oroxylum indicum* Cocoon characters; Silk Filament Characters and the Electrophoretic patterns of esterase activity of silk worm *Bombyx mori* (L.) (Race: PM x CSR2). *Research Journal of Recent Sciences* Vol. 3(IVC-2014), 1-5 (2014) ISSN 2277-2502 . www.isca.in, www.isca
64. Vitthalrao B. Khyade; and Jiwan P. Sarawade (2012). Contents of protein and activity of protease and amylase in the mid gut homogenate of fifth instar larvae of *Bombyx mori* L. (PM x CSR2) fed with herbal drug (Kho-Go) treated mulberry leaves. *International Journal of Science and Nature* Vol.3 (3): 526 – 530 www.scienceandnature.org ISSN 2229 – 6441.
65. Vitthalrao B. Khyade; Kajal P. Shukla and Jeevan P. Sarawade (2012). Juvenile Hormone activity of some non mulberry plant extractives through inhibition of chitin deposition in the integument of fifth instar larvae of silk worm, *Bombyx mori* (L) (Race: PM x CSR2). *Research Journal of Recent Sciences*, Vol. 1 (Issue:ISC-2112): 1-6. www.isca.in ISSN 2277 – 2502.
66. Vitthalrao B. Khyade; Karel Slama; Rajendra D. Pawar and Sanjay V. Deshmukh (2015). Influence of Various Concentrations of Acetone Solution of Retinol on Pattern of Chitin Deposition in the Integument of Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (PM X CSR2). *Journal of Applicable Chemistry*. 2015, 4 (5): 1434 – 1445. www.joac.info
67. Vitthalrao B. Khyade; Karel Slama; Rajendra D. Pawar and Sanjay V. Deshmukh (2015). Influence of Various Concentrations of Acetone Solution of Retinol on Pattern of Chitin Deposition in the Integument of Fifth Instar Larvae of Silkworm, *Bombyx mori* (L) (PM X CSR2). *Journal of Medicinal Plants Studies*. Volume 3 Issue 5 Part C: 124 – 131. <http://www.plantsjournal.com/archives/?year=2015&vol=3&issue=5&part=C>
68. Vitthalrao B. Khyade; Poonam M. Patil; Kalyani R. Jaybhay; Rasika G. Gaikwad; Ganga V. Mhamane; Vivekanand V. Khyade; Kavita H. Nimbalkar and Sneha G. Jagtap (2007). Effect of digoxin on economic parameters of silk worm, *Bombyx mori* (L). *Journal of Zoological Society of India: Bioinformatics*: 23 – 31. (Editors: B. N. Pandey; Sadhana Deshpande; A. K. Tripathi and A. D. Adsool) (Publisher: A P H Publishing Corporation, New Delhi) (ISBN 13: 9788131302200 / ISBN 10: 8131302202).
69. Vitthalrao B. Khyade; Poonam M. Patil; Kalyani R. Jaybhay; Rasika G. Gaikwad; Ganga V. Mhamane; Vivekanand V. Khyade; Kavita H. Nimbalkar and Sneha G. Jagtap (2007). Effect of digoxin on mid gut glucosidase activity in silkworm, *Bombyx mori* (L). *Journal of Zoological Society of India: Bioinformatics*: 32 – 48. (Editors: B. N. Pandey; Sadhana Deshpande; A. K. Tripathi and A. D. Adsool) (Publisher: A P H Publishing Corporation, New Delhi) (ISBN 13: 9788131302200 / ISBN 10: 8131302202).
70. Vitthalrao B. Khyade; Poonam M. Patil; Sharad G. Jagtap; Sunanda V. Khyade and Jeevan P. Sarawade (2010). Effect of Methanolic Extractives of Roots of *Achyranthus aspera* on Larval Body Wall Chitin in the Fifth Instars of Silkworm, *Bombyx mori* (L) (Race: PM x CSR₂). *Advances in Plant Sciences*. 23(I): 309 313.

71. Vitthalrao B. Khyade; Sakharam B. Patil; Sunanda V. Khyade and Ganesh P. Bhawane (2002). Influence of Acetone maceratives of *Vitis vinifera* on larval parameters of silkworm, *Bombyx mori* (L). Indian Journal of Comparative Animal Physiology Vol. 21 (1): 14 – 18.
72. Vitthalrao B. Khyade; Sakharam B. Patil; Sunanda V. Khyade and Ganesh P. Bhawane (2003). Influence of Acetone maceratives of *Vitis vinefera* on economic parameters of silkworm *Bombyx mori* (L). Indian Journal of Comparative Animal Physiology Vol. 21 (1): 28 – 32.
73. Vitthalrao B. Khyade; Sunanda V. Khyade and Vivekanand V. Khyade (2009). Influence of mealy bug infestation on mulberry leaves on the silkworm, *Bombyx mori*(L). Eco friendly Insect Pest Management: 325 – 328. The book edited by Dr. S. Iganacimuthu Director, Entomology Research Institute, Loyola College, Chennai – 600034. ISBN: 81 – 88901 – 37 – 7 . Publisher: Elite Publishing House Pvt. Ltd. New Delhi.
74. Vitthalrao B. Khyade; Uma S. Ghantaloo and Vandana D. Shinde (2007). Various effects of anti-biotics on selected parameters of silkworm *Bombyx mori*(L). Journal of Zoological Society of India: Bioinformatics: 11 – 22. (Editors: B. N. Pandey; Sadhana Deshpande; A. K. Tripathi and A. D. Adsool) (Publisher: A P H Publishing Corporation, New Delhi) (ISBN 13: 9788131302200 / ISBN 10: 8131302202).
75. Vitthalrao B. Khyade; Vivekanand V. Khyade and Rhidim D. Mote (2014). Influence of Acetone extractive of *Oroxylum indicum* (L) on cocoon characters, silk filament character and electrophoretic patterns of esterase activity of silkworm, *Bombyx mori* (L) (Race: PM x CSR2). Recent Trends in Zoology(Pages: 12-22). Editor: Dr. R. K. Kasar; Publisher: Dr. L. S. Matkar Principal, New Arts, Commerce and Science College, Shevgaon Dist. Ahmednagar – 414502 (M.S.) India. ISBN: 978-93-84916-68-8.
76. Vitthalrao B. Khyade; Vrushali D. Shinde and Shraddha S. Maske (2016). Influence of the diterpenoids (Retinol and Phytol) (Race: PM x CSR2) on the cocoon and silk parameters in silkworm, *Bombyx mori* (L) (Race: PM x CSR2). World Scientific news 42 (2016): 1-12. www.worldscientificnews.com .
77. Vitthalrao B.Khyade; Vivekanand V. Khyade and Randy Wayne Schekman (2015). Utilization of the topical application of Limonene to the fifth instar larvae of the silkworm, *Bombyx mori* (L) (Race: PM X CSR2) for the parameters of Larvae, Cocoon and Silk filament. International Journal of Bioassay 4 (02): 3632 – 3635.ISSN: 2278-778X www.ijbio.com
78. Vitthalrao Khyade, Edvard Moser and May – Britt Moser (2015). INFLUENCE OF AQUEOUS MACERATIVES OF SEED POWDER OF SYZIGIUM CUMINI (L) ON THE MID GUT ENZYME ACTIVITY IN THE FIFTH INSTAR LARVAE OF SILK WORM, *BOMBYX MORI* (L) (Race: PM x CSR2). World Journal of Pharmaceutical Research Volume 4, Issue 6:997 – 1008. (ISSN 2277– 7105).www.wjpr.net
79. Waldbauer, G.P., 1968. The consumption and utilization of food by insects. Adv. Insect Physiol., 5: 229-288.
80. Rahul Pratapsing Patil; Saniya Shahanwaz Patel and Vitthalrao Bhimasha Khyade (2018). Assesment of Antlion, Myrmeleon Species for The Economic Optimization in Malegaon Sheti farm of Krishi Vidnyan Kendra Baramati India. International Academic Journal of Economics Vol. 5, No. 2, 2018, pp. 67-78. ISSN 2454-2474 www.iaiest.com
81. Vitthalrao B. Khyade and Rajashri Vivekanand Khyade (2018). Strengthening Role of Information and Communication Technology in Global Society. International Academic Journal of Accounting and Financial Management Vol. 5, No. 2, 2018, pp. 42-49. ISSN 2454-2350 www.iaiest.com
82. Vitthalrao B. Khyade and Manfred Eigen (2018). Key Role of Statistics for the Fortification of Concepts in Agricultural Studies. International Academic Journal of Innovative Research Vol. 5, No. 3, 2018, pp. 32-46. ISSN 2454-390X www.iaiest.com

83. Vitthalrao B. Khyade and Sidney Altman (2018). Use of Herbal Terpenoid for topical application to fifth instars of silkworm, *Bombyx mori* (L). International Academic Journal of Science and Engineering Vol. 5, No. 3, 2018, pp. 43-62. ISSN 2454-3896 www.iaiest.com
84. Vitthalrao Bhimasha Khyade; Shubhangi Shankar Pawar and Jiwan Pandurang Sarwade (2018). Novel Sacrificial Medicinal Repositories: Halfa grass, *Desmostachya bipinnata* (L.) and Cogon grass, *Imperata cylindrica* (L.). World Scientific News WSN 100 (2018) 35-50 EISSN 2392-2192. www.worldscientificnews.com
85. Vitthalrao B. Khyade (2018). World Day for Cultural Diversity for Dialogue and Development: 21 May. International Academic Journal of Innovative Research Vol. 5, No. 3, 2018, pp. 47-57. ISSN 2454-390X www.iaiest.com
86. Vitthalrao B. Khyade (2018). Globalization: Necessary Evil for the Qualitative Society International Academic Journal of Science and Engineering. Vol. 5, No. 3, 2018, pp. 76-97. ISSN 2454-3896 www.iaiest.com
87. Vitthalrao B. Khyade (2018). Influence of Leaf Decoction of Mulberry, *Morus alba* (L.) on Streptozotocin Induced Diabetes in Brown Rat, *Rattus norvegicus* (L.). International Journal of Research in Science and Engineering Vol. 6, No. 3, 2018, pp. 1-23. ISSN 2347-9353 www.scientificrc.com
88. Vitthalrao B. Khyade (2018). Herbals and Their Compounds Targeting Pancreatic Beta Cells for the Treatment of Diabetes. International Journal of Scientific Studies Vol. 6, No. 3, 2018, pp. 1-44. ISSN 2348-3008 www.scientificrc.com
89. Vitthalrao Bhimasha Khyade and Jiwan Pandurang Sarwade (2018). Novel Sacrificial Medicinal Repositories: Halfa grass, *Desmostachya bipinnata* (L.) and Cogon grass, *Imperata cylindrica* (L.). International Journal of Research in Science and Engineering Vol. 6, No. 3, 2018, pp. 24-36. ISSN 2347-9353 www.scientificrc.com
90. Vitthalrao B. Khyade (2018). Bacterial diversity in the alimentary canal of earthworms. Journal of Bacteriology & Mycology 2018;6(3):183–185. DOI: 10.15406/jbmoa.2018.06.00200 <http://medcraveonline.com/JBMOA/JBMOA-06-00200.pdf>
91. Vitthalrao B. Khyade (2018). The levels of plasma glucose and insulin; oxidative stress and body weight in streptozotocin induced diabetic rats treated with aqueous solution of Moracin. International Journal of Research in Science and Engineering Vol. 6, No. 3, 2018, pp. 37-57. ISSN 2347-9353 www.scientificrc.com
92. Vitthalrao B. Khyade and Peeyush M. Pahade (2018). Utilization of Aqueous Solution of Sericin from the Silk Cocoons of Silkworm, *Bombyx mori* (L.) For the Control of Diabetes in Brown Rat, *Rattus norvegicus* (L.). International Journal of Scientific Studies Vol. 6, No. 3, 2018, pp. 82-100. ISSN 2348-3008 www.scientificrc.com
93. Vitthalrao Bhimasha Khyade (2018). The Mulberry, *Morus alba* (L.): The Marvelous Entity of Herbal Formulation for Wealthy Human Health. International Journal of Scientific Studies Vol. 6, No. 3, 2018, pp. 58-81. ISSN 2348-3008 www.scientificrc.com
94. Sharad G. Jagtap and Vitthalrao B. Khyade (2018). Oxidative Stress Reducing Capabilities of Moracin, the Novel Compound from the Fruits of Mulberry, *Morus alba* (L) (AH927). Asian Journal of Science and Technology Vol. 09, Issue, 05, pp.8126-8133, May, 2018. <http://www.journalajst.com>
95. Vitthalrao Bhimasha Khyade (2018). The Influence of Sericin (separately); Moracin (separately); Sericin and Moracin (both together) on blood glucose level; body weight and water consumption in the non-diabetic and streptozotocin-induced diabetic rats. *Journal of research in health science*. Vol. 1, No. 3, 2018, pp. 58-84. DOI 10.26739/2523-1243 www.journalofresearch.org

96. Vitthalrao Bhimasha Khyade and Shinya Yamanaka (2018). Sericin from the cocoons of Silkworm, *Antheraea Mylitta* (L) and *Bombyx Mori* (L) for the Reduction in Hydrogen Peroxide Induced Oxidative Stress in Feline Fibroblasts. *International Journal of Scientific Research in Chemistry (IJSRCH)* | Online ISSN: 2456-8457 © 2018 IJSRCH | Volume 3 | Issue 4: 01-16. <http://ijsrch.com/archive.php?v=3&i=6&pyear=2018>
97. Manali Ramesh Rao Shinde, Seema Karna Dongare, Vitthalrao Bhimasha Khyade (2018). Qualitative Silk Cocoons in Silkworm, *Bombyx Mori* (L) Through the Topical Application of Acetone Macerative of Powder of *Ganoderma* Fruiting Body And Acetone Solution of It's Triterpenoid (Lucidone –D). *International Journal of Scientific Research in Chemistry (IJSRCH)* | Online ISSN: 2456-8457 © 2018 IJSRCH | Volume 3 | Issue 4: 17 – 34. <http://ijsrch.com/archive.php?v=3&i=6&pyear=2018>
98. Seema Karna Dongare, Manali Rameshrao Shinde, Vitthalrao Bhimasha Khyade (2018). Mathematical Inverse Function (Equation) For Enzyme Kinetics. *International Journal of Scientific Research in Chemistry (IJSRCH)* | Online ISSN: 2456-8457 © 2018 IJSRCH | Volume 3 | Issue 4: 35– 42. <http://ijsrch.com/archive.php?v=3&i=6&pyear=2018>
99. Kajal Appasaheb Pondkule, Manisha Mahendra Nalwade, Vitthalrao B. Khyade (2018). Changes in the Mid gut enzyme activity in the fifth instar larvae of silkworm, *Bombyx mori* (L) fed with mulberry leaves treated with aqueous solution of seed powder of *Syzigium cumini* (L) (Race: Bivoltine Cross Breed [(CSR6 x CSR26) x CSR2 x CSR27]). *International Journal of Scientific Research in Chemistry (IJSRCH)* | Online ISSN: 2456-8457 © 2018 IJSRCH | Volume 3 | Issue 4:43– 52. <http://ijsrch.com/archive.php?v=3&i=6&pyear=2018>
100. Amruta Chandrakant Nimbalkar, Samiksha Sunil Pisal, Mansi Ramesh Das, Vitthalrao B. Khyade (2018). Bioconversion of Garbage: Garden Waste (GW); Kitchen Waste (KW) and Combination of Both Garbage: Garden Waste and Kitchen (GW +KW) into Vermicompost through the use of earthworm, *Eisenia fetida* (L). *International Journal of Scientific Research in Chemistry (IJSRCH)* | Online ISSN: 2456-8457 © 2018 IJSRCH | Volume 3 | Issue 4:53– 67. <http://ijsrch.com/archive.php?v=3&i=6&pyear=2018>
101. Manisha Mahendra Nalwade, Kajal Appasaheb Pondkule, Vitthalrao B. Khyade (2018). The reflection of feeding the mulberry leaves treated with water solution of seed powder of *Syzigium cumini* (L) into Profiles of Protein (Total) in the fifth instar larvae of silk worm, *Bombyx mori* (L) Race - bivoltine, crossbreed: [(CSR6 x CSR26)] x [CSR2 x CSR27]). *International Journal of Scientific Research in Chemistry (IJSRCH)* | Online ISSN: 2456-8457 © 2018 IJSRCH | Volume 3 | Issue 4:68– 76. <http://ijsrch.com/archive.php?v=3&i=6&pyear=2018>
102. Wheeler, D.E. and Nijhout, H.F. (1983). Soldier determination in the ant, *Pheidole bicarinata*: Hormonal control of caste and size within castes. *J. Insect Physiol.* 29: 847-854.
103. Wheeler, D.E. and Nijhout, H.F. (2003) A perspective for understanding the modes of juvenile hormone action as a lipid signaling system. *Bio Essay*, 25:994–1001.
104. Wyatt, G.R. and Davey, K.G. (1996) Cellular and molecular actions of juvenile hormone. II. Roles of juvenile hormone in adult insects. *Adv. Insect Physiol.* 26:1–155.
105. Xu, Ran; Fazio, Gia C.; Matsuda, Seiichi P.T. (February 2004). "On the origins of triterpenoid skeletal diversity". *Phytochemistry.* 65 (3): 261–291. doi:10.1016/j.phytochem.2003.11.014.
106. Zera, A.J and Tiebel, K.C. (1988). Brachypterizing effect of group rearing, juvenile hormone-III, and methoprene on winglength development in the wing-dimorphic cricket, *Gryllus rubens*. *J. Insect Physiol.* 34:489–498.
107. Zera, A.J and Zhao, Z.(2004) Effect of a juvenile hormone analogue on lipid metabolism in a wingpolymorphic cricket: Implications for the endocrine-biochemical bases of life-history trade-offs. *Papers in the Biological Sciences, University of Nebraska – Lincoln.* Posted at DigitalCommons@University of Nebraska - Lincoln.